

NEOLIGNANS FROM *URBANODENDRON VERRUCOSUM**

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Abstract—Branch wood of the shrub *Urbanodendron verrucosum* from the Atlantic forest of southern Brazil contains the benzofuranoid neolignans licarin-A, licarin-D, porosin and the novel porosin-B, as well as the tetrahydrofurane neolignans austrobailignan-7, calopiptin and the novel verrucosin. The acid-catalysed rearrangement of the porosins yields the corresponding dihydrofutoenones.

INTRODUCTION

The monotypical genus *Urbanodendron* (Lauraceae) is represented by the shrub *U. verrucosum* (Nees) Mez which occurs in the Atlantic forest region of Rio de Janeiro State. The branch wood contains the benzofurane neolignans licarin-A (1a)[2] and licarin-D (1b)[3], the tetrahydrobenzofurane neolignans porosin (2a)[4] and porosin-B (2b), as well as the tetrahydrofurane neolignans austrobailignan-7 (3a)[5], calopiptin (3b)[6, 7] and verrucosin (3c).

For reasons stated in a previous paper of this series[8] nomenclature and numbering of neolignans follow the rules outlined in a recent review[9].

RESULTS AND DISCUSSION

The novel compounds 2b and 3c are substitutional variants of known co-specific isolates and the elucidation of their structures relied on trivial spectral comparisons. Both the previously known porosin (2a) and the new porosin-B (2b) not only give comparable ¹H NMR spectra (Table 1), but also identical ORD curves. Thus not only do they possess the same relative configurations but also the same absolute ones. The porosins were submitted to treatment with acid (in methanol), in order to provide additional evidence for the *cis*-relationship between the Ar-7 and the Me-8 (¹H NMR δ 0.5) groups[4]. It was hoped that acid-catalysed opening and closure of the benzylic C–O bond might induce the formation of porosins in which an Ar-7/Me-8 *trans*-relationship would be characterized by ¹H NMR methyl signals at lower field (*ca* δ 1.1[2]). Indeed, ¹H NMR evidence showed the reaction products to include *ca* 1:1 mixtures of 2a+2c and 2b+2d respectively. The separation of the epimers from these mixtures proved

difficult and was not completed. Furthermore, the reaction gave methanol addition products of the porosins in a yield of 35% in addition to the spiro-derivatives 4a and 4b.

Compound 4b has been obtained previously by the catalytic hydrogenation of futoenone (4c), a neolignan from *Piper futokadzura* Sieb. et Zucc.[10]. All reported data for dihydrofutoenone are identical with the data obtained for the acid rearrangement product of porosin-B. With respect to the ¹H NMR data, excepting only the signals due to different aromatic substitution, this fact applies also to 4a, the rearrangement product of porosin. Both rearrangement products therefore have identical configurations.

A topic which merits discussion concerns the location of the double bond in 4a, 4b. Although this could, *a priori*, also be placed between C-5' and C-6', such an alternative can be dismissed. Indeed, compounds 2a, 2b, 4a and 4b must have identical chromophores, since their UV spectra are so similar [λ_{\max} nm: 259 ± 1 , 288 ± 2 inf. (ϵ 15 800, 4000)]. An α,β -unsaturated ketone with an α -ether substituent would give a UV maximum at *ca* 268 nm with a substantially smaller extinction coefficient[4].

The porosin (2a, 2b)–dihydrofutoenone (4a, 4b) rearrangement is mechanistically analogous to the known burchellin (5)–guianin (6)–futoenone (4c) interconversions[11, 12].

The structure of verrucosin (3c) was established by comparison with the synthetic racemate[13] (superimposable ¹H NMR data) and by preparation of a dimethyl ether (3d) (identical in all respects, including $[\alpha]_D$, to veraguensin[14]).

EXPERIMENTAL

Isolation of constituents. Plant material, identified by Professor Klaus Kubitzki, Hamburg University, was collected at Tijuca Forest, Rio de Janeiro. Dried and powdered branch wood (1.5 kg) was percolated with C₆H₆. The extract (15 g) was submitted to CC (300 g Si gel). The following fractions were eluted with the indicated solvents: A–D

*Part LXVII in the series "The Chemistry of Brazilian Lauraceae. For Part LXVI see ref. [1]. Based in part on the M.Sc. Thesis presented by A. de F. D., on leave of absence from Universidade Federal da Paraíba, to Universidade de São Paulo (1980).

Table 1. ^1H NMR spectral data of neolignans*

	2a [4]		2b	4a		4b [10]
H-2	6.76 <i>d</i> (2)	}				
H-5	6.83 <i>dd</i> (8, 2)		6.5–6.8	6.6–6.9	<i>m</i>	6.6–6.9
H-6	6.93 <i>d</i> (8)					
H-7	5.89	<i>d</i> (5)	5.74	}	×	×
H-8	2.6	<i>m</i>	2.4–2.7			
3H-9	0.52	<i>d</i> (7)	0.50	0.75	<i>d</i> (6)	0.73
H-3'	5.59	<i>s</i>	5.44	5.63	<i>s</i>	5.63
H-5'	4.02	<i>dd</i> (12, 5)	3.92	3.68	<i>m</i>	3.69
H-6'	2.22	<i>dd</i> (12, 5)	2.26	2.27		2.27
H-6'	1.92	<i>t</i> (12)	1.85	2.14		2.14
H-7'	2.69 <i>dd</i> (14, 7)	}				
H-7'	2.36 <i>dd</i> (14, 7)		2.4–2.6	×		×
H-8'	5.9–6.1	<i>m</i>	5.6–6.2	4.93	<i>m</i>	4.95
2H-9'	5.3–5.4	<i>m</i>	5.1–5.3	×		×
MeO-5'	3.62	<i>s</i>	3.53	3.58	<i>s</i>	3.58
2MeOAr	3.90 <i>s</i>	—	—	3.90	<i>s</i>	—
CH ₂ O ₂ Ar	—	—	5.87 <i>s</i>	—	<i>s</i>	5.95

*Chemical shifts in δ from internal TMS for CDCl₃ solutions at 220 MHz (2a) and 60 MHz (all other compounds); coupling constants (Hz) in brackets. $\times \delta$ 1.7–2.5, *m* 8H.

(C₆H₆), E (C₆H₆–CH₂Cl₂, 19 : 1), F (C₆H₆–CH₂Cl₂, 9 : 1), G (C₆H₆–CH₂Cl₂, 1 : 1), H (CH₂Cl₂–MeOH, 99 : 1), I (CH₂Cl₂–MeOH, 19 : 1). A (0.2 g) consisted of aliphatic esters. B (1 g) was a mixture of terpenes. C (0.6 g) was re-crystallized from MeOH to 1a (100 mg). The mother liquor was re-chromatographed on a Si gel column. Elution with C₆H₆ gave terpenes, 1b (50 mg) and 3a (50 mg). D (2 g) was re-chromatographed on a Si gel column. Elution with C₆H₆–CH₂Cl₂ mixtures of gradually increasing polarity gave 3a (130 mg) and 3b (300 mg). E (0.5 g) was a mixture of 3a, 3b and 3c. F (0.2 g) was purified by prep. TLC (Si gel) to 3c (50 mg). G (0.2 g) was a mixture of 3c and 2b. H (1 g) was crystallized from MeOH–H₂O to 2b (750 mg). I (1 g) was purified by prep.–TLC (Si gel, Et₂O) to 2a (800 mg).

Identification of known compounds. The known compounds were identified either by direct comparison with authentic samples (1a, 1b, 2a) or by comparison of mp, $[\alpha]_D$ and spectral data with published data (3a, 3b).

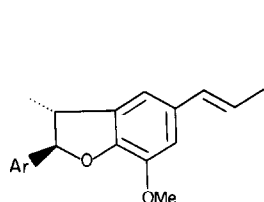
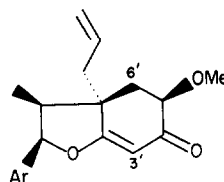
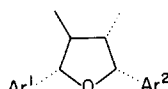
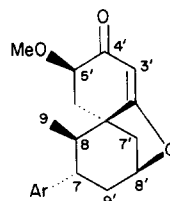
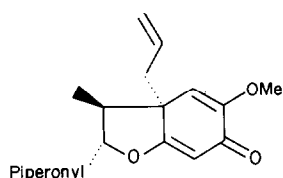
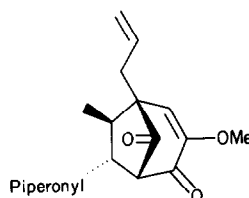
Rel-(7R, 8S, 1'R, 5'R)- Δ^8 -5'-methoxy-3, 4-methylenedioxy-1', 4', 5', 6'-tetrahydro-4'-oxo-7.O.2', 8.1'-neolignan (porosin-B, 2b). Mp 114–116° (MeOH–H₂O). (Found: 342.1448; C₂₀H₂₂O₅ requires 342.1467). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 260, 290 inf. (ϵ 14 000, 5100). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1660, 1630, 1500, 1450. ORD (*c* 1 mg/10 ml MeOH): $[\theta]_{335}^D + 6800$, $[\theta]_{315}^D$ 0, $[\theta]_{280}^D - 41\,000$, $[\theta]_{262}^D$ 0, $[\theta]_{252}^D + 85\,500$. MS *m/z* (rel. int.): 343 [*M* + 1]⁺ (10), 342 [*M*]⁺ (50), 312 (69), 284 (26), 269 (22), 256 (21), 241 (20), 215 (33), 175 (100), 173 (25), 162 (78), 161 (23), 150 (20), 149 (68), 135 (100), 115 (53), 103 (70), 91 (90).

(7R, 8S, 7'S, 8'S)-4, 4'-Dihydroxy-3, 3'-dimethoxy-7.O.7', 8.8'-neolignan (verrucosin, 3c). Oil (Found: 344.1585; C₂₀H₂₄O₅ requires: 344.1624). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 280 (ϵ 17 100). IR $\nu_{\text{max}}^{\text{film}}$ cm⁻¹: 3350, 1600, 1500. ^1H NMR (60 MHz, CDCl₃) δ 6.7–7.1 (*m*, 6 ArH), 4.28 (*d*, *J* = 8.5 Hz, H-7), 5.01 (*d*, *J* = 8 Hz, H-7'), 1.5–2.5 (*m*, H-8, H-8'), 0.63 (*d*, *J* = 6.5 Hz, 3H-9), 1.05 (*d*, *J* = 6 Hz, 3H-9'), 3.81 and 3.87 (2*s*, 2 MeO). $[\alpha]_D^{25} + 38^\circ$ (CHCl₃; *c* 0.25). **Dimethyl ether** (3d) (3c, CH₂N₂, Et₂O), mp and lit. [6, 14] mp 128–129°, $[\alpha]_D^{25}$ and lit. [6, 14] $[\alpha]_D + 35^\circ$ (CHCl₃; *c* 0.19).

Porosin-dihydrofutoenone rearrangement. Porosin (2a, 100 mg) and TsOH (40 mg) were maintained in MeOH (15 ml) at 50° (8 hr). The solvent was evaporated under vacuum and the residue separated by prep. TLC (Si gel, Et₂O) into a mixture of 2a + 2c (30 mg), an addition product of 2a and MeOH (25 mg) and 4a (35 mg). Porosin-B (2b) treated in the same way gave 2b + 2d, an addition product of 2b and MeOH and 4b in similar proportions.

Rel-(7S, 8R, 1'S, 5'R, 8'S)-2', 8'-epoxy-3, 4, 5'-trimethoxy-1', 4', 5', 6'-tetrahydro-4'-oxo-7.9', 8.1'-neolignan (4a). Mp 162–164°. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 259, 285 inf. (ϵ 15 800, 4000). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1660, 1640, 1510. ORD (*c* 1 mg/10 ml MeOH): $[\theta]_{258}^D - 20\,000$, $[\theta]_{307}^D$ 0, $[\theta]_{275}^D + 67\,300$.

Rel-(7S, 8R, 1'S, 5'R, 8'S)-2', 8'-epoxy-5'-methoxy-3, 4-methylenedioxy-1', 4', 5', 6'-tetrahydro-4'-oxo-7.9', 8.1'-neolignan (4b). Mp and lit. [10] mp 171–173°. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 259, 288 inf. (ϵ 15 800, 4000). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1660, 1640, 1490. ORD (*c* 1 mg/10 ml MeOH): $[\theta]_{277}^D - 20\,500$, $[\theta]_{308}^D$ 0, $[\theta]_{275}^D + 70\,000$.

**1a** Ar = Guaiacyl**1b** Ar = Veratryl**2a** Ar = β -Veratryl**2b** Ar = β -Piperonyl**2c** Ar = α -Veratryl**2d** Ar = α -Piperonyl**3a** Ar¹ = Piperonyl, Ar² = Guaiacyl**3b** Ar¹ = Piperonyl, Ar² = Veratryl**3c** Ar¹ = Ar² = Guaiacyl**3d** Ar¹ = Ar² = Veratryl**4a** Ar = Veratryl**4b** Ar = Piperonyl**4c** Ar = Piperonyl, $\Delta^{5',6'}$ **5****6**

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REFERENCES

- Gomes, M. C. C. P., Yoshida, M. and Gottlieb, O. R. *Phytochemistry* (in press).
- Aiba, C. J., Corrêa, R. C. and Gottlieb, O. R. (1973) *Phytochemistry* **12**, 1163.
- Fernandes, J. B., Gottlieb, O. R. and Maia, J. G. S. (1976) *Phytochemistry* **15**, 1033.
- Aiba, C. J., Gottlieb, O. R., Yoshida, M., Mourão, J. C. and Gottlieb, H. E. (1976) *Phytochemistry* **15**, 1031.
- Murphy, S. T., Ritchie, E. and Taylor, W. (1975) *Aust. J. Chem.* **28**, 81.
- McAlpine, J. B., Riggs, N. V. and Gordon, P. G. (1968) *Aust. J. Chem.* **21**, 2095.
- Doskotch, R. W. and Flom, M. S. (1972) *Tetrahedron* **28**, 4711.
- Díaz, D., P. P., Yoshida, M. and Gottlieb, O. R. (1980) *Phytochemistry* **19**, 285.
- Gottlieb, O. R. (1978) *Progr. Chem. Org. Nat. Prod.* **35**, 1.
- Ogiso, A., Kurabayashi, M., Takahashi, S., Mishima, H. and Woods, M. C. (1970) *Chem. Pharm. Bull.* **18**, 105.
- Büchi, G. and Mak, C.-P. (1977) *J. Am. Chem. Soc.* **99**, 8073.
- de Alvarenga, M. A., Brocksom, U., Gottlieb, O. R. and Yoshida, M. (1978) *J. Chem. Soc. Chem. Commun.* 831.
- Sarkanen, K. V. and Wallis, A. F. A. (1973) *J. Chem. Soc. Perkin Trans. 1* 1869.
- Crossley, N. S. and Djerassi, C. (1962) *J. Chem. Soc.* 1459.